Lytic Phage for Foodborne Pathogens Detection Iryna Sorokulova<sup>1</sup>, Eric Olsen<sup>2</sup>, Ludmila Globa<sup>1</sup>, James Barbaree<sup>3</sup>, Vitaly Vodyanoy<sup>1</sup> <sup>1</sup>Department of Anatomy, Physiology and Pharmacology, College of Veterinary Medicine, Auburn University, Auburn, AL 36849; <sup>2</sup>Clinical Research Laboratory, 81st Medical Group, Keesler AFB MS USA 39534; <sup>3</sup>Department of Biological Sciences, n University, Auburn, AL 36849

Foodborne pathogens are among the most significant problems in maintaining the health of the population. The leading causes of foodborne illnesses in the United States are *Salmonella* and *Shigella* (1). *Staphylococcus aureus* is among top five pathogens contributing to domestically acquired foodborne illnesses. Since foodborne infections have a dramatic impact on morbidity and mortality, particularly of infants and children, timely detection of these pathogens is highly important. One of the promising approaches for detection of pathogenic bacteria in environment is the use of lytic phage boisensors.

We isolated lytic phage against *Staphylococcus aureus* with wide spectrum of hosts, including MRSA (Fig.1 A). Comparison of isolated phage's lytic activity with activity of *S. aureus* phage from the American Type Culture Collection showed that only isolated phage 12600 was effective against all tested MRSA strains (Fig.1 B).

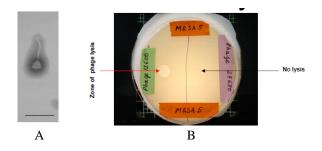


Figure 1. *S. aureus* phage: A – Transmission electron micrographs of isolated phage (bar – 200 nm); B – phage 12600 lytic activity against MRSA (left); phage 27690 has no lytic activity (right)

Co-cultivation of phage 12600 with MRSA resulted in quick lysis of bacterial culture (Fig. 2A). This phage, adsorbed on the gold surface remains its lytic activity (Fig. 2B).

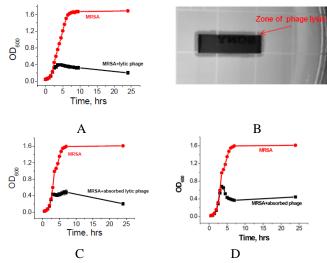


Figure 2. Lysis of MRSA by *S. aureus* phage 12600: A- co-cultivation with free phage; B and C – lysis by gold-absorbed phage; D – lysis by absorbed phage after storage

Co-cultivation of MRSA strain with adsorbed phage also resulted in lysis of bacterial culture. The kinetic of bacterial lysis was the same as with free phage (Fig. 2C). Immobilized phage remains alive and active after 6 days of storage at 4°C (Fig. 2D). Obtained results show that adsorbed phage can recognize, capture and lyse MRSA cells. This observation opens perspectives for fabrication of lytic phage biosensors for detection of pathogenic bacteria.

Next step in our research was isolation of phage for detection of *Salmonella* and *Shigella* pathogens. Newly isolated phage had unique spectrum of lytic activity – it was effective against all 23 tested strains of *Salmonella* spp., 2 strains of *Shigella* spp. and showed no activity against other closely related bacteria (Fig. 3; Table 1).

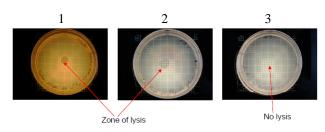


Figure 3. Selectivity of *Salmonella* phage: 1 - *Salmonella typhimurium* DT 104 2- *Shigella flexneri;* 3 - *Yersinia enterocolotica* 

Bacterial cultures,	Bacterial cultures,
sensitive to	resistant to <i>Salmonella</i>
<i>Salmonella</i> phage	phage
<ol> <li>S. typhimurium Health 9491</li> <li>S. typhimurium DT 104 Dairy</li> <li>S. diarisonae</li> <li>S. ganama SA 3583</li> <li>S. Jindica SA 4401</li> <li>S. derby SARB 10</li> <li>S. trybhimurium LT2</li> <li>S. trybhimurium IT3</li> <li>S. trybhimurium G787</li> <li>S. typhimurium 6787</li> <li>S. trybhimurium Health 1390</li> <li>S. S. Jopon SA 4910</li> <li>S. stame SA 41106</li> <li>S. styphimurium S20-96</li> <li>S. typhimurium DT 104 Swine</li> <li>S. typhimurium ATCC 13311</li> <li>S. typhimurium ATCC 13311</li> <li>S. typkimurium S05</li> </ol>	<ol> <li>Pseudomonas aeruginosa</li> <li>Pseudomonas fluorescens</li> <li>Eschericha coli ATCC 11775</li> <li>Kiebsiella pneumoniae 13882</li> <li>Yersinia enterocolitica</li> <li>Proteus mirabilis</li> <li>Staphylococcus aureus ATCC 12600</li> <li>S. aureus ATCC 27690</li> <li>S. aureus 10292</li> <li>S. aureus 10292</li> <li>S. aureus 10497</li> <li>S. aureus 10497</li> <li>S. aureus MRSA 1</li> <li>S. aureus MRSA 5</li> <li>S. aureus MRSA 5</li> <li>S. aureus MRSA 45</li> <li>S. aureus MRSA 45</li> <li>B. aureus MRSA 45</li> </ol>

Table 1. Bacterial test-cultures for detection ofSalmonella phage selectivity

Strong bacterial specificity of isolated phages makes it possible to create biosensors with near-to-realtime capacity in recognition of foodborne pathogens.

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## **REFERENCES:**

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