

A Review of Single-Cell Manipulation Techniques for Microfluidic Lab-on-a-Chip Systems

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Cell behavior is traditionally inferred from in vitro bulk measurements of large heterogeneous populations, yielding ensemble averages that obfuscate probes of discrete and intrinsic cellular properties¹⁻⁴. Conversely, single-cell diagnostic methods directly probe each cell as an individual unto itself^{1-2,4}, facilitating finer studies of viability⁵⁻⁶, pathogenesis⁷⁻⁸, cell mechanics⁹⁻¹¹, links between biochemistry and cellular behavior¹²⁻¹⁴, surface redox activity¹⁵, cellular hydrodynamics¹⁶, etc. Single-cell diagnostics also afford the quantification of population heterogeneity and facilitate the identification, characterization, and separation of subpopulations¹⁷⁻¹⁸. The most ubiquitous single-cell analysis method is flow cytometry (FC), which yields high-throughput, multiparameter analyses of individually probed cells¹⁹⁻²⁰ - facilitating identification of homogeneous subpopulations and studies of links between single- and multi-cell behaviour^{4,21-22}. However, FC does not afford the tracking, manipulation, dynamic analysis, and subsequent retrieval of specific individual cells²³. The other conventional single-cell diagnostic techniques (automated microscopy²⁴, laser scanning cytometry²⁵, capillary electrophoresis²⁶, and laser capture microdissection²⁷) are low-throughput methods ill-suited to single-cell assays²³.

Advances in polymeric micro- and nano-fabrication afford increasingly sophisticated single-cell diagnostic tools^{3,28-30}. In particular, microfluidic (MF) technologies offer several advantages over traditional tools, including: reduced material and power consumption; faster processing; portability; greater potential parallelization and integration; and lower fabrication costs^{19,30}. Such MF technologies feature prominently in integrated lab-on-a-chip (LoC) and micro-total analysis system (μ TAS) platforms^{19,30}. Advances in LoC/ μ TAS technology promise high-throughput chips capable of tracking, manipulating, analyzing, and retrieving specific individual cells-satisfying applications beyond the capabilities of traditional techniques. As an example: our lab has been working towards an integrated LoC/ μ TAS device intended to trap cells in an array of physical single-cell traps, perform an immunobioassay identifying the cells secreting the antibodies of interest, and selectively retrieve those cells to create monoclonal cell lines producing said antibodies in large quantities³¹⁻³².

The first step in developing such a LoC/ μ TAS device is to design a subsystem to manipulate individual cells, preferably to guide and hold them in arrays of reversible single-cell traps. This paper is primarily a critical review of advances in single-cell manipulation, relevant to such a LoC/ μ TAS platform. A wide variety of techniques will be examined, including: the magnetic trapping of labelled cells³³⁻³⁷; the manipulation of cells using optical tweezers³⁸⁻⁴²; the polarization and attraction/repulsion of cells via dielectrophoresis⁴³⁻⁴⁶; the manipulation of cells using the electromagnetic fields of surface plasmons⁴⁷⁻⁴⁸; and the trapping of cells in arrays of micro-scale wells^{23,31-32,49-50}, perfusive hydrodynamic traps^{30,51-53}, and microvortexes⁵⁴⁻⁵⁵. The potential for various single-cell analyses and the subsequent retrieval of cells afforded by each method shall be addressed.

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